Electromagnetic signatures of the preclinical and prodromal stages of Alzheimer’s disease

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Biomarkers useful for the predementia stages of Alzheimer’s disease are needed. Electroencephalography and magnetoencephalography (MEG) are expected to provide potential biomarker candidates for evaluating the predementia stages of Alzheimer’s disease. However, the physiological relevance of EEG/MEG signal changes and their role in pathophysiological processes such as amyloid-β deposition and neurodegeneration need to be elucidated. We evaluated 28 individuals with mild cognitive impairment and 38 cognitively normal individuals, all of whom were further classified into amyloid-β-positive mild cognitive impairment (n = 17, mean age 74.7 ± 5.4 years, nine males), amyloid-β-negative mild cognitive impairment (n = 11, mean age 73.8 ± 8.8 years, eight males), amyloid-β-positive cognitively normal (n = 13, mean age 71.8 ± 4.4 years, seven males), and amyloid-β-negative cognitively normal (n = 25, mean age 72.5 ± 3.4 years, 11 males) individuals using Pittsburgh compound B-PET. We measured resting state MEG for 5 min with the eyes closed, and investigated regional spectral patterns of MEG signals using atlas-based region of interest analysis. Then, the relevance of the regional spectral patterns and their associations with pathophysiological backgrounds were analysed by integrating information from Pittsburgh compound B-PET, fluorodeoxyglucose-PET, structural MRI, and cognitive tests. The results demonstrated that regional spectral patterns of resting state activity could be separated into several types of MEG signatures as follows: (i) the effects of amyloid-β deposition were expressed as the alpha band power augmentation in medial frontal areas; (ii) the delta band power increase in the same region was associated with disease progression within the Alzheimer’s disease continuum and was correlated with entorhinal atrophy and an Alzheimer’s disease-like regional decrease in glucose metabolism; and (iii) the global theta power augmentation, which was previously considered to be an Alzheimer’s disease-related EEG/MEG signature, was associated with general cognitive decline and hippocampal atrophy, but was not specific to Alzheimer’s disease because these changes could be observed in the absence of amyloid-β deposition. The results suggest that these MEG signatures may be useful as unique biomarkers for the predementia stages of Alzheimer’s disease.

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Introduction

Alzheimer’s disease is the most common neurocognitive disorder, with an estimated prevalence of ~60–70% of the 47.5 million people with dementia worldwide (http://www.who.int/mediacentre/factsheets/fs362/en/). Recent disease-modifying clinical trials for Alzheimer’s disease (Sperling et al., 2014; Sevigny et al., 2016) have emphasized the importance of early intervention in the preclinical stages, which are categorized as mild cognitive impairment (MCI) and preclinical Alzheimer’s disease stages in the diagnostic criteria of the National Institute on Aging and the Alzheimer’s Association (NIA-AA) (Albert et al., 2011; McKhann et al., 2011; Sperling et al., 2011). Therefore, the registration of individuals with preclinical stages in global platforms aiming to facilitate clinical trials such as the global Alzheimer’s platform (Cummings et al., 2016) and the European Prevention of Alzheimer’s Dementia (Ritchie et al., 2016) has become the most recent global trend. However, as MCI manifests as a heterogeneous clinical status in which the clinical outcome is considerably variable (Larrieu et al., 2002), identification of MCI individuals with biomarker evidence of Alzheimer’s disease is crucial (Jack et al., 2016; Wolz et al., 2016). Such individuals are termed ‘MCI due to Alzheimer’s disease’ (Albert et al., 2011) by the NIA-AA criteria, or ‘prodromal Alzheimer’s disease’ by the International Working Group on Alzheimer’s disease (Dubois et al., 2010, 2014). Biomarker information is more crucial to identify individuals in the preclinical stages of Alzheimer’s disease who are cognitively asymptomatic but who also have evidence of amyloid-β deposition in the brain.

Biomarkers for Alzheimer’s disease are important not only for identifying these high-risk individuals, but also for assessing the disease status or understanding the pathophysiological processes of disease progression. Amyloid-β deposition is the earliest pathognomonic signature of Alzheimer’s disease, which starts decades before the actual onset of Alzheimer’s disease (Morris, 2005; Bateman et al., 2012; Villemagne et al., 2013). Thus, biomarkers for amyloid-β deposition, such as amyloid-PET imaging signatures or decreased amyloid-β$_{1-42}$ and amyloid-β$_{1-42}$/amyloid-β$_{1-40}$ ratio in the CSF, are considered to be the most ‘upstream’ markers in the pathological cascade of Alzheimer’s disease (Jack et al., 2013). However, amyloid-β deposition does not necessarily represent progression to Alzheimer’s disease, as many subjects with abundant amyloid-β deposition are able to live their natural lifespan cognitively intact (Snowdon, 1997). Therefore, identification of downstream biomarkers that can act as surrogate markers of disease progression is also important. These markers include the CSF concentrations of total tau and phosphorylated tau (Blennow et al., 2010), tau-PET imaging markers (Maruyama et al., 2013; Harada et al., 2016), reduced glucose metabolism predominantly in the posterior cingulate, precuneus, and tempo-parietal cortices as measured by fluorodeoxyglucose (FDG)-PET (Drzezga et al., 2003; Anchisi et al., 2005; Mosconi, 2005), brain atrophy in the medial temporal area as assessed by structural MRI (Risacher et al., 2009; Chételat et al., 2012; Doré et al., 2013), and accelerated cognitive decline (Storandt et al., 2009; Lim et al., 2014b).

EEG and magnetoencephalography (MEG) are expected to be useful for providing unique biomarker candidates (Stomrud et al., 2010; Fernández et al., 2013; López et al., 2016), as they are direct measures of primary neural activity and have very fine temporal resolution (in the order of milliseconds). Moreover, their non-invasive nature allows for repeated measurements to monitor the disease status or to evaluate the effects of intervention. Patients with Alzheimer’s disease and MCI generally show slowing of oscillatory brain activity (Stam, 2010; Lizio et al., 2011; López et al., 2014b; Engels et al., 2016), and this slowing is associated with several factors such as cognitive status, risk of progression to dementia (Fernández et al., 2006b; Prichep, 2007; López et al., 2016), hippocampal atrophy (Fernández et al., 2003), hypometabolism (Rodriguez et al., 1998), CSF tau levels (Jelic et al., 1998; Stomrud et al., 2010; Kramberger et al., 2013), APOE4 genotype (Lehtovirta et al., 1996; de Waal et al., 2013; Cuesta et al., 2014), and low cholinergic activity (Riekkinen and Sirviö, 1990). However, the usefulness of EEG/MEG characteristics as biomarkers for the evaluation of preclinical stages of Alzheimer’s disease is not yet fully established. One main reason is that only a few studies of preclinical stages of Alzheimer’s disease have been performed in which EEG/MEG was combined with amyloid-β biomarker information (Jelic et al., 1998; Stomrud et al., 2010; Kramberger et al., 2013; Gouw et al.,
Therefore, the main objective of this study was to explore potential electrophysiological signatures that may act as surrogate markers of the pathophysiological changes that occur in the predementia stages of Alzheimer’s disease. Further, we investigated the relevance of these biomarker candidates to their pathophysiological backgrounds by combining MEG data with amyloid-β biomarker data obtained using Pittsburgh compound B (PiB)-PET, downstream markers for neurodegeneration obtained using FDG-PET, and structural MRI.

Accordingly, we analysed regional spectral patterns of resting state MEG signals in 17 amyloid-β-positive individuals with MCI (MCIp), 11 amyloid-β-negative individuals with MCI (MCIn), 13 amyloid-β-positive cognitively normal (CNp) individuals, and 25 amyloid-β-negative CN individuals (CNn). To understand the characteristics of potential MEG signatures, we first explored the effects of amyloid-β deposition (amyloid-β positive versus amyloid-β negative) and clinical status (MCI versus CN), and their interaction by means of a two-way design. Second, we performed group-wise comparisons to further extract group-specific characteristics. Further, relationships between the MEG power markers and pathophysiological processes, including cognitive decline, regional glucose metabolism, and grey matter volume, were analysed. Through these assessments, we separately identified MEG signatures that were (i) related to amyloid-β deposition; (ii) related to downstream pathophysiological changes within the Alzheimer’s disease continuum; and (iii) non-specific changes related to general cognitive decline or neurodegeneration.

**Materials and methods**

**Participants**

This investigation was a part of the Multimodal Neuroimaging for Alzheimer’s disease Diagnosis (MULNIAD) study, which is a prospective longitudinal study targeting normal ageing, MCI, and Alzheimer’s disease that was conducted at the National Center for Geriatrics and Gerontology (NCGG) in Obu, Japan. All participants were native Japanese individuals who were recruited from among community-dwelling aged individuals or outpatients at the National Hospital for Geriatric Medicine, NCGG. The study was approved by the Ethics Committee of NCGG, and all participants provided written informed consent. The original sample comprised 33 patients (2017; Nakamura et al., 2017). Therefore, the main objective of this study was to explore potential electrophysiological signatures that may act as surrogate markers of the pathophysiological changes that occur in the predementia stages of Alzheimer’s disease. Further, we investigated the relevance of these biomarker candidates to their pathophysiological backgrounds by combining MEG data with amyloid-β biomarker data obtained using Pittsburgh compound B (PiB)-PET, downstream markers for neurodegeneration obtained using FDG-PET, and structural MRI.

All participants underwent a comprehensive battery of neuropsychological tests and neuroimaging assessments including PiB-PET, FDG-PET, structural MRI, and MEG. All examinations were carried out within ~1 month of each other.

**PiB-PET**

**Image acquisition**

3D PET imaging for 50–70 min after intravenous injection of 555 ± 185 MBq 11C-PiB was carried out using a PET CT camera, Biograph True V (Siemens Healthcare). X-ray CT was performed before PET imaging for attenuation correction.

**Visual rating and classification**

Visual rating of PiB-PET images was conducted according to the previously described protocol (Kaneko et al., 2014), which followed the method reported by Rabinovici et al. (2011). This was used for classification of participants into amyloid-β-positive (CNp and MCIp) and amyloid-β-negative (CNn and MCIn) groups (Supplementary material).

**Quantitative image analysis**

Standardized uptake value ratio (SUVR) images were generated individually using the Automated Anatomical Labeling (AAL) atlas (Tzourio-Mazoyer et al., 2002). As the representative measure of the quantitative amyloid-β burden, the mean cortical PiB-SUVR was obtained by averaging the SUVRs of the frontal, parietal, and temporal regions of interest from the AAL atlas. The PiB-SUVR images were spatially smoothed using a Gaussian kernel filter with a full-width at half-maximum of 8 mm. Whole-brain voxel-wise regression analysis for MEG power data was performed with these smoothed images using the Statistical Parametric Mapping (SPM8, Wellcome Trust Centre for Neuroimaging, University College, London, UK) software suite (Supplementary material).

**FDG-PET**

**Image acquisition**

Using the same scanner and attenuation correction method as used for PiB-PET, 18F-FDG-PET images were obtained. Whole-brain voxel-wise regression analysis for the MEG power data with the FDG-PET images was performed using SPM8 (Supplementary material).

**MRI**

**Image acquisition**

High-resolution 3D T1-weighted images were acquired using a Trio 3 T scanner (Siemens) and used for volumetric analysis (Supplementary material). T2-weighted and fluid attenuated inversion recovery images were also acquired to assess brain lesions.
Table 1  Participant demographics

<table>
<thead>
<tr>
<th></th>
<th>CNp (n = 13)</th>
<th>CNn (n = 25)</th>
<th>MCIp (n = 17)</th>
<th>MCIin (n = 11)</th>
<th>Statistics (P-value)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex (M / F)</td>
<td>7/6</td>
<td>11/14</td>
<td>9/8</td>
<td>8/3</td>
<td>0.47</td>
</tr>
<tr>
<td>Age (y)</td>
<td>71.8 ± 4.4</td>
<td>72.5 ± 3.4</td>
<td>74.7 ± 5.4</td>
<td>73.8 ± 8.8</td>
<td>0.43</td>
</tr>
<tr>
<td>Education (y)</td>
<td>12 ± 3</td>
<td>12 ± 3</td>
<td>11 ± 3</td>
<td>12 ± 3</td>
<td>0.59</td>
</tr>
<tr>
<td>MMSE</td>
<td>28.77 ± 1.09</td>
<td>28.60 ± 1.38</td>
<td>26.29 ± 1.65</td>
<td>26.64 ± 2.87</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>ADAS-cog</td>
<td>5.74 ± 2.19</td>
<td>5.87 ± 2.67</td>
<td>9.09 ± 2.04</td>
<td>9.35 ± 3.28</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>LM1</td>
<td>20.77 ± 7.25</td>
<td>20.36 ± 6.40</td>
<td>11.47 ± 4.99</td>
<td>12.00 ± 6.26</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>LM2</td>
<td>16.23 ± 6.85</td>
<td>16.24 ± 6.17</td>
<td>4.18 ± 5.14</td>
<td>7.00 ± 7.20</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>CDR</td>
<td>0</td>
<td>0</td>
<td>0.5</td>
<td>0.5</td>
<td>-</td>
</tr>
<tr>
<td>CDR-SOB</td>
<td>0.04 ± 0.14</td>
<td>0.08 ± 0.19</td>
<td>1.85 ± 1.03</td>
<td>1.73 ± 1.19</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>GDS</td>
<td>2.00 ± 1.29</td>
<td>2.08 ± 1.63</td>
<td>2.41 ± 1.66</td>
<td>2.82 ± 1.66</td>
<td>0.58</td>
</tr>
<tr>
<td>APOE-4 (%)</td>
<td>4/13 (30.8)</td>
<td>4/25 (16.0)</td>
<td>1/17 (76.5)</td>
<td>0/11 (0.0)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>PiB-mcSUVR</td>
<td>1.44 ± 0.19</td>
<td>1.13 ± 0.06</td>
<td>1.89 ± 0.25</td>
<td>1.13 ± 0.06</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>VSRAD score</td>
<td>0.54 ± 0.18</td>
<td>0.69 ± 0.42</td>
<td>1.34 ± 0.62</td>
<td>1.14 ± 0.67</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

Values are presented as mean ± SD. Statistical analyses were performed using the chi square test (sex, APOE-4) and one-way ANOVA (others). ADAS-cog = Alzheimer’s Disease Assessment Scale-Cognitive Component-Japanese version; APOE-4 = positive for apolipoprotein E-4; CDR = Clinical Dementia Rating; CDR-SOB = Sum of Boxes of CDR; GDS = Geriatric Depression Scale; LM1/LM2 = Logical Memory III from the Wechsler Memory Scale–Revised (paragraphs A and B); MMSE = Mini-Mental Status Examination; PiB-mcSUVR = mean cortical SUVR of PiB-PET; VSRAD score = the degree of grey matter atrophy of the medial temporal region using a z-score computed by VSRAD (also see Supplementary Fig. 1).

Volumetric analysis

Atrophy in the medial temporal region, including the hippocampus, head to tail of the hippocampus, and amygdala, was quantitatively assessed using the Voxel-based Specific Regional Analysis System for Alzheimer’s Disease (VSRAD® advance, Eisai Co., Ltd., Tokyo, Japan) software, which is based on SPM8 and Diffeomorphic Anatomical Registration Through Exponentiated Lie Algebra (DARTEL; Matsuda et al., 2012; Matsuda, 2013) (Supplementary material).

The spatially normalized and grey matter-segmented images created by VSRAD were also used for whole-brain voxel-wise regression analysis using SPM8, and the associations between the MEG power values and grey matter volume were estimated.

MEG

Data acquisition

The MEG measurements were performed using a 306-channel whole-head MEG system (Vectorview, ElektaNeuramag) located in a magnetically shielded room at the NCGG. Participants sat comfortably on a chair with their eyes closed, and resting state MEG signals were measured for 5 min with a sampling rate of 1000 Hz (online bandpass anti-alias filtering at 0.1–330 Hz) (Supplementary material). The arousal level of each subject was monitored with a video camera (WV-CL934, Panasonic) and also checked via a conversation immediately following the measurement session. If a subject reported feeling sleepy during the session, we gave him/her sufficient time to feel more awake and performed the measurement again.

Computation of the power spectra

After data preprocessing (Supplementary material), at least 20 artefact-free fragments (trials) of continuous 4-s MEG signals (80 s of brain activity) were obtained from all participants, and 20 of these clean trials were randomly selected from each subject to equalize the number of trials. The time series was filtered using a broadband filter (1.4–55 Hz) with a Finite Impulse Response filter (order, 1500) designed with a Hannings window.

Using the realistic single-shell model with a 1-cm spacing grid (2455 nodes), source reconstruction was performed using a Linearly Constrained Minimum Variance Beamformer (Van Veen et al., 1997) (Supplementary material).

The MEG power spectra of each node were computed for all artefact-free trials. A frequency-of-interest range from 1.5–55 Hz (in 0.5-Hz steps) was used. To obtain the average frequency content of each trial, we applied a multitaper method with discrete prolate spheroidal sequences as windowing functions and 1 Hz smoothing. Trials were averaged across subjects, obtaining a matrix with dimension: 2455 nodes × 108 frequency steps × 66 subjects. For each node, the relative power was calculated by normalizing with the total power over the 1.5- to 55-Hz range (Jelic et al., 2000).

Design for data analyses

To separately detect MEG power spectral changes related to amyloid-β deposition and clinical status, the data were analysed in a two-way design that tested for the main effects of amyloid-β deposition (amyloid-β-positive groups versus amyloid-β-negative groups), clinical status (MCI groups versus CN groups), and their interaction. Also, group-wise comparisons were conducted to test for the effects of amyloid-β deposition within the CN groups (CNp versus CNn), and within the MCI groups (MCIp versus MCIin). In addition, the effects of clinical status were tested within the amyloid-β-positive groups (CNp versus MCIp), and within the amyloid-β-negative groups (CNn versus MCIin). It is well known that ageing strongly affects the MEG/EEG power spectrum (Rossini et al., 2007), and thus, the analyses were conducted while adjusting for the effects of age using analysis of covariance (ANCOVA).

Region of interest-based analysis

The source-reconstructed MEG power data were first analysed by atlas-based region of interest analysis to visualize the
characteristics of the regional spectral patterns as waveforms. We set 10 representative regions of interest by referring to the AAL atlas (Tzourio-Mazoyer et al., 2002), so that the whole cortical mantle was roughly covered. Five regions of interest were related to the default mode network (Buckner et al., 2008), because amyloid-β first accumulates in areas associated with the default mode network (Mintun et al., 2006), and glucose hypometabolism in the posterior hub of the default mode network is an established marker for disease progression (Small et al., 2000). Accordingly, the 10 regions of interest were: frontal medial cortex, anterior cingulate cortex, left and right frontal cortices, left and right temporal cortices, left and right inferior parietal lobules, precuneus/posterior cingulate cortex, and occipital cortex (Supplementary Table 1).

For these regions of interest, the effects of amyloid-β deposition and clinical status were visualized in a spatio-frequent domain by plotting the F-values, which were computed by two-way and group-wise ANCOVAs for each frequency step (0.5 Hz) while adjusting for the effects of age.

Node-based whole-brain analysis

The region of interest-based analyses were performed mainly for visualization purposes, whereas actual statistical analyses were conducted on node-based whole-brain analysis.

The source template with 2455 nodes in a 1-cm spacing grid was segmented into 72 AAL (Tzourio-Mazoyer et al., 2002) regions of interest that included all AAL regions of interest but excluded the cerebellum, basal ganglia, thalamus, amygdala, insula, and olfactory cortices using the AAL version made for SPM8 included in Fieldtrip software (Oostenveld et al., 2011). The 72 regions of interest included 1137 of the original 2455 nodes. The source-reconstructed MEG power data were analysed with a matrix of 1137 nodes × 108 frequency steps × 66 subjects.

To overview spatio-frequential topography of the effects of amyloid-β deposition and clinical status, F-values for the two-way and group-wise ANCOVAs were computed at each of the 1137 nodes for each frequency and overlaid on the standard brain of the Montreal Neurological Institute.

Significant clusters, which showed significant effects of amyloid-β deposition or clinical status, were explored using cluster-based permutation tests (CBPT) (Maris and Oostenveld, 2007) performed over the 1137 nodes, using Fieldtrip toolbox (Oostenveld et al., 2011). To enhance the frequential resolution, the results of the above spatio-frequential F-value maps as well as the region of interest-based analyses were used to define specific frequency ranges of interest for each candidate. Within each classical frequency band [i.e. delta (2–4 Hz), theta (4–8 Hz), and alpha (8–12 Hz)], we defined three frequency steps bands centred at the frequency with the largest effect. Then, whole-brain power spectral data were averaged within the corresponding frequency band and submitted to the CBPT. The CBPT were executed with 10 000 repetitions to create a null distribution for each comparison. This null distribution was obtained by shuffling the original values and performing a two-way or one-way ANCOVA test with age as a confounding covariate for testing the two-way or group-wise comparisons, respectively. The maximum statistic at each repetition was kept for the permutation distribution. The CBPT P-value represents the proportion of the permutation distribution with F-values greater than or equal to the F-value of the original data. The alpha level was set to 0.05 for the CBPT P-value. To enhance the spatial resolution of the results, the critical value for the clusters configuration was fixed to 0.01. Only those clusters that were retained after the CBPT were used for subsequent analyses as potential ‘MEG power markers’. Power values of all nodes included in a cluster were averaged individually and used as power marker values for the subsequent correlation, regression, or receiver operating characteristic (ROC) analyses.

Correlations between the MEG power markers and the mean cortical PiB-SUVR values, VSRAD scores, or the scores for the cognitive tests were assessed by multiple correlation analysis using age as a confounding covariate.

The ROC analyses were used to evaluate the performances of MEG power markers to predict amyloid-β positivity or negativity (Supplementary material). ROC analyses were performed, adjusting for the effects of age as follows: for each MEG power marker, a predictive formula that included age as a confounder was built by using binomial logistic regression analysis. Then, the predictive values were computed and used for the age-adjusted ROC analyses.

Statistical analyses were performed using Matlab R2009b (The Mathworks Inc., Natick, MA, USA) and SPSS v. 21 (IBM, Armonk, NY, USA) software. All tests were two-tailed, and the significance level threshold was set at P < 0.05 unless explicitly stated otherwise.

Results

Participant demographics

The demographic characteristics of the participants are shown in Table 1. No differences were present in sex, age, or educational level among the four groups. Overall, performance on the neuropsychological tests was significantly lower in the MCI groups than in the CN groups, except for the Geriatric Depression Scale score. The proportion of APOE-ε4 carriers was significantly higher in the MCIp group compared to all the other groups. The mean cortical SUVR of PiB-PET values was significantly higher in the MCIp group compared to all the other groups, and the CNp group showed a higher mean cortical PiB-SUVR value than the MCIp and CNn groups. The VSRAD scores were significantly higher (implying a higher level of atrophy) in the MCIp group than in the CNp group (P = 0.019), and in the MCIp group than in the CNp group (P < 0.001; Supplementary Fig. 1).

General profile of the regional power spectra

To provide an overview of the power-frequency profile of the MEG resting state signals for each group, the relative power spectra and their subtractions of two-way comparisons were plotted at 10 representative regions of interest (Fig. 1A). In the frequency range below 7 Hz, which corresponds to the classical description of theta and delta bands, the MCI groups generally showed larger power values than the CN groups in all regions of interest.
Main effects of clinical status were found in the prefrontal regions of interest (frontal medial cortex and anterior cingulate cortex) (Fig. 1A, orange lines). In particular, the CNp group showed the strongest alpha power among all four groups in these regions of interest, whereas the MCIn group was the weakest. In addition, the alpha power in these regions of interest was weaker in the MCI groups than the CN groups.

### MEG signatures showing effects of amyloid-β deposition and clinical status

To visualize the effects of amyloid-β deposition and clinical status in a spatio-frequency domain, F-values of the two-way and group-wise ANCOVAs at each of the representative regions of interest were plotted (Fig. 1B and C). In the two-way analyses, stronger main effects of amyloid-β deposition were found in the prefrontal regions of interest (frontal medial cortex and anterior cingulate cortex) at a frequency range peaking around 9 Hz (Fig. 1B, bottom left). In contrast, stronger main effects of clinical status were found in most regions of interest in a frequency range lower than 7 Hz (Fig. 1B, bottom right). In addition, significant main effects of clinical status were found in the prefrontal regions of interest at a peak frequency ~9 Hz. We found no prominent interaction between amyloid-β deposition and clinical status (Fig. 1B, top). In the group-wise analyses for effects of amyloid-β deposition, both within the CN groups (CNp versus CNn) and within the MCI groups (MCIp versus MCIn), comparisons showed prominent effects in the prefrontal regions of interest at peak frequencies of 10 Hz (Fig. 1C, top left) and 9 Hz (Fig. 1C, bottom left), respectively. For the group-wise effects of clinical status, the spatio-frequency profiles appeared quite different when comparing the amyloid-β-positive groups (MCIp versus CNp) and the amyloid-β-negative groups (MCIn versus CNn). Within the amyloid-β-negative group comparisons, the spatio-frequency
Table 2 Significant MEG power markers

<table>
<thead>
<tr>
<th>Comparisons</th>
<th>Regions</th>
<th>Classical band</th>
<th>Peak frequency</th>
<th>Power (SD) Aβp</th>
<th>Power (SD) Aβn</th>
<th>ANCOVA F value</th>
<th>p-value*</th>
<th>η²p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Effects of amyloid-β deposition</td>
<td></td>
<td></td>
<td></td>
<td>Aβp</td>
<td>Aβn</td>
<td></td>
<td></td>
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<tr>
<td>Main effects</td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group effects</td>
<td>CNp &gt; CNn</td>
<td>Prefrontal</td>
<td>Alpha</td>
<td>0.036(0.008)</td>
<td>0.032(0.006)</td>
<td>11.325</td>
<td>0.001</td>
<td>0.157</td>
</tr>
<tr>
<td>Within CN</td>
<td>CNp &gt; CNn</td>
<td>Prefrontal</td>
<td>Alpha</td>
<td>0.029(0.006)</td>
<td>0.023(0.005)</td>
<td>10.821</td>
<td>0.002</td>
<td>0.236</td>
</tr>
<tr>
<td>Within MCI</td>
<td>MCIp &gt; MCIn</td>
<td>Prefrontal</td>
<td>Alpha</td>
<td>0.037(0.007)</td>
<td>0.028(0.005)</td>
<td>13.318</td>
<td>0.001</td>
<td>0.348</td>
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<tr>
<td>Effects of clinical status</td>
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<tr>
<td>Main effects</td>
<td>MCI &gt; CN</td>
<td>Posterior</td>
<td>Delta</td>
<td>0.019(0.004)</td>
<td>0.017(0.003)</td>
<td>12.340</td>
<td>0.001</td>
<td>0.168</td>
</tr>
<tr>
<td>Group effects</td>
<td>MCIn &gt; MCIn</td>
<td>Occipitotemporal</td>
<td>Delta</td>
<td>0.019(0.005)</td>
<td>0.016(0.003)</td>
<td>18.473</td>
<td>0.000</td>
<td>0.359</td>
</tr>
<tr>
<td>Within Aβp</td>
<td>MCIp &gt; CNp</td>
<td>Occipital</td>
<td>Delta</td>
<td>0.021(0.003)</td>
<td>0.017(0.004)</td>
<td>10.757</td>
<td>0.003</td>
<td>0.285</td>
</tr>
<tr>
<td></td>
<td>MCIp &gt; CNp</td>
<td>Prefrontal</td>
<td>Delta</td>
<td>0.027(0.005)</td>
<td>0.022(0.004)</td>
<td>11.278</td>
<td>0.002</td>
<td>0.295</td>
</tr>
</tbody>
</table>

*Statistical test using ANCOVA adjusted for age.
Aβn/p = amyloid-β-negative/positive.

Figure 2 MEG power markers representing the effects of amyloid-β (Aβ) deposition and clinical status. The shape of each cluster is overlapped on the standard brain of the Montreal Neurological Institute. Each arrow indicates the peak frequency where the maximum effect was detected. The red colour indicates that the amyloid-β-positive groups showed larger power than the amyloid-β-negative groups, the green colour indicates larger power in the MCI groups than the CN groups, and the blue colour indicates the opposite.
profile was similar to that observed in the two-way analyses, showing stronger effects at peak frequencies around 5–6 Hz in all regions of interest and stronger effects around 9 Hz in the frontal regions of interest (Fig. 1C, top right). In contrast, the amyloid-β-positive group comparisons demonstrated stronger effects of the clinical status in the prefrontal and occipital regions of interest at peak frequencies around 2–4 Hz (Fig. 1C, bottom right).

These region of interest-based findings were confirmed with whole-brain analyses that computed F-values for each node involved in the whole cortical regions. Topographical maps of F-values at each frequency, which are shown in Supplementary Fig. 2A–C, demonstrated similar spatio-frequency profiles as the region of interest-based analyses for each effect related to amyloid-β deposition or clinical status.

Finally, we extracted statistically significant clusters that showed significant effects of amyloid-β deposition or clinical status, using the CBPT (Maris and Oostenveld, 2007) applied for the node-level whole-brain analysis. These clusters are listed in Table 2, and were used for the subsequent analyses as potential MEG power markers. Accordingly, we identified 12 MEG power markers as shown in Fig. 2. Three markers represented the effects of amyloid-β deposition as alpha band power augmentation in the amyloid-β-positive groups compared with the amyloid-β-negative groups in the prefrontal regions. The centre frequencies were 9.5 Hz for the main effects, and 10.5 Hz and 9 Hz for the within-group effects in the CN and MCI groups, respectively (Fig. 2, red clusters). The other nine power markers represented the effects of clinical status as either a power increase in the MCI groups (MCI > CN) within the delta and theta bands (Fig. 2, green clusters), or an alpha power decrease in the MCI groups (Fig. 2, blue clusters). The power markers for the group effects within the amyloid-β-negative groups were similar to those observed in the main effects, and appeared to be a type of subset of the main effects. On the other hand, two power markers for the group effects of clinical status within the amyloid-β-positive groups were represented as a delta band power increase in the MCIp group in the occipital and medial prefrontal regions. Detailed anatomical correspondence between these MEG power marker clusters and the regions of interest of the AAL atlas are shown in Supplementary Table 2.

### Links between MEG power markers and pathophysiological processes

#### Amyloid-β deposition

To assess the relevance of the MEG power markers to pathophysiological processes, including amyloid-β deposition, neurodegeneration, and cognitive decline, multiple correlation analyses between the MEG power marker values and cognitive/imaging scores were performed while adjusting for the effects of age (Table 3).

The three power markers that represented the effects of amyloid-β deposition as prefrontal alpha power augmentation did not show any significant correlations with cognitive scores

### Table 3 Correlations between the MEG power markers and cognitive/imaging scores

<table>
<thead>
<tr>
<th>Regions</th>
<th>Classical band</th>
<th>Peak frequency Hz</th>
<th>Partial correlation (r) adjusted for age</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>mcSUVR</td>
</tr>
<tr>
<td><strong>Effect of amyloid-β deposition</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Main effects</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Prefrontal</td>
<td>Alpha</td>
<td>9.5</td>
<td>0.138</td>
</tr>
<tr>
<td>Group effects</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Within CN</td>
<td>Prefrontal</td>
<td>Alpha</td>
<td>10.5</td>
</tr>
<tr>
<td>Within MCI</td>
<td>Prefrontal</td>
<td>Alpha</td>
<td>9</td>
</tr>
<tr>
<td><strong>Effect of clinical status</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Main effects</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Posterior</td>
<td>Delta</td>
<td>2.5</td>
<td>0.150</td>
</tr>
<tr>
<td>Global</td>
<td>Theta</td>
<td>4.5</td>
<td>0.227</td>
</tr>
<tr>
<td>Prefrontal</td>
<td>Alpha</td>
<td>9.5</td>
<td>0.115</td>
</tr>
<tr>
<td>Group effects</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Within A/ln</td>
<td>Occipitotemporal Delta</td>
<td>2.5</td>
<td>−0.023</td>
</tr>
<tr>
<td></td>
<td>Occipitotemporal Theta</td>
<td>4.5</td>
<td>0.136</td>
</tr>
<tr>
<td></td>
<td>Prefrontal Delta</td>
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<td>0.180</td>
</tr>
<tr>
<td></td>
<td>Prefrontal Alpha</td>
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<td>−0.027</td>
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<tr>
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<td>Occipital Delta</td>
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<td>0.188</td>
</tr>
<tr>
<td></td>
<td>Prefrontal Delta</td>
<td>3.5</td>
<td>0.323</td>
</tr>
</tbody>
</table>

Asterisks indicate statistically significant correlations: (*P < 0.05, **P < 0.01, ***P < 0.001). A/ln/p = amyloid-β-negative/positive ADAS-Jcog = Alzheimer’s Disease Assessment Scale-Cognitive Component-Japanese version; APOE=ε4 = positive for apolipoprotein E ε4; CDR = Clinical Dementia Rating; CDR-SOB = Sum of Boxes of CDR; GDS = Geriatric Depression Scale; LM1/LM2 = Logical Memory I/II from the Wechsler Memory Scale–Revised (paragraphs A and B); MMSE = Mini-Mental State Examination; PiB-mcSUVR = mean cortical SUVR of PiB-PET; VSRAD score = the degree of grey matter atrophy of the medial temporal region using a z-score computed by VSRAD.
such as the Mini-Mental State Examination, Alzheimer’s Disease Assessment Scale-Cognitive Component-Japanese version (ADAS-Jcog), Logical Memory I/II from the Wechsler Memory Scale–Revised (LM1, LM2), and Clinical Dementia Rating-Sum of Boxes (CDR-SOB) scores, or with the quantitative measures of medial temporal atrophy as assessed with VSRAD scores (Table 3). Among them, however, two power markers showed group effects within CN and within MCI and demonstrated significant positive correlations with the mean cortical PiB-SUVR values with a partial $r = 0.412$ ($P = 0.011$) and $0.468$ ($P = 0.014$), respectively (Table 3, mean cortical SUVR). The correlation coefficients became higher when the correlations were analysed against local PiB-SUVR values that were computed when limited within each power marker cluster. The partial $r$ values for the within-CN and within-MCI markers were $0.517$ ($P = 0.001$) and $0.502$ ($P = 0.008$), respectively (Fig. 3, top). The results suggested that these power marker values are associated more
with local amyloid-β deposition than with global amyloid-β burden. In addition, correlations were also analysed with only the amyloid-β-positive groups (CNp and MCIp). Within the CNp group, the dose-dependent effect of local amyloid-β load on the alpha power augmentation was immediately high (partial $r = 0.527$), although it did not reach significance ($P = 0.079$), whereas such an effect was not observed within the MCIp group (partial $r = -0.059$, $P = 0.828$).

To elucidate the topographical relationship between these power markers and cerebral amyloid-β deposition further, we conducted whole-brain regression analyses using PiB-SUVR images. The results demonstrated that the power marker values were significantly associated with amyloid-β deposition, mainly in the prefrontal regions (Fig. 3, middle). In particular, the within-CN power marker showed a close topographical relationship in the anterior cingulate and medial prefrontal cortices with local amyloid-β deposition (Fig. 3, bottom), suggesting that the link between the alpha power augmentation and local amyloid-β load in these regions is stronger in the CN groups compared with that in the MCI groups.

**Neurodegeneration**

In contrast, the nine MEG power markers, which represented the main effects or group effects of clinical status, were not significantly correlated with the mean cortical PiB-SUVR values (Table 3). However, all of them demonstrated significant correlations with at least one cognitive score, including the ADAS-Jcog, LM1, LM2, or CDR-SOB (Table 3). In particular, the low-frequency power markers (delta and theta) generally showed stronger correlations than the alpha-range power markers. All these low-frequency markers, except the within-amyloid-β-positive occipital delta marker, also demonstrated a significantly positive correlation with the VSRAD scores (Table 3), indicating that these power markers are associated with the degree of medial temporal atrophy.

To investigate the relevance of the power markers to neurodegenerative processes further, we conducted whole-brain voxel-wise regression analyses between the power marker values and regional grey matter volume using individual structural MRIs. Also, regression analyses between the power values and regional glucose metabolism were conducted using individual FDG-PET images. The power markers, which represented the main effects of clinical category in the posterior delta power (2.5 Hz) and global theta power (4.5 Hz), demonstrated that their power values were significantly negatively correlated with the grey matter volume in the hippocampal region [cluster level family wise error (FWE)-corrected $P < 0.05$] (Fig. 4A). Because the analyses were conducted based on whole-brain voxel-based morphometry, the results indicated that the relationship to the structural change was very specifically limited to the hippocampal volume. The prefrontal alpha power (9.5 Hz) did not show such correlations. The regression analyses of these power markers with FDG-PET images did not show any significant clusters.

Similarly, the power markers, which represented the group effects of clinical category within the amyloid-β-negative groups in the occipitotemporal delta (2.5 Hz) and theta (4.5 Hz) power, also demonstrated significant negative correlations with the hippocampal grey matter volume (Fig. 4B), whereas the prefrontal theta (5.5 Hz) and alpha (9 Hz) power did not show such correlations. These power markers did not show any significant associations with FDG-PET images.

Notably, the power marker, which represented the group effects of clinical category within the amyloid-β-positive groups in the prefrontal delta (3.5 Hz), demonstrated significantly negative correlations with the grey matter volume mainly in the entorhinal cortex (Fig. 4C). Further, the regression analysis of this marker with FDG-PET images showed a significant negative correlation with regional glucose metabolism in the precuneus and posterior cingulate cortices (Fig. 4D), indicating that increased medial prefrontal delta power was associated with Alzheimer’s disease-type hypometabolism. The power marker in the occipital delta (3 Hz) did not show such relationships.

**Discussion**

In this study, we extracted several MEG power markers that represented the either the effects of amyloid-β deposition or the effects of the clinical status. Further, this study disclosed the relevance of these power markers to pathophysiologival processes, including amyloid-β deposition, neurodegeneration, and cognitive decline in detail. To the best of our knowledge, this is the first report that systematically identifies the regional spectral patterns of the spontaneous electromagnetic brain activity in MCI and CN subjects by combining MEG with multiple imaging modalities, including PiB-PET, FDG-PET, and structural MRI.

**Effects of amyloid-β deposition on regional power spectra**

The results demonstrated that the effects of amyloid-β deposition were manifested as the alpha power increment in prefrontal regions. This alpha augmentation was significantly correlated with amyloid-β burden in the same or adjacent prefrontal regions, and this topographical association was more evident within the CN group than within the MCI group. Several studies have shown that individuals in the predementia stages of Alzheimer’s disease exhibit functional upregulation in the frontal areas of the brain. Using MEG, our group reported that functional connectivity between the anterior cingulate cortex and temporo-occipital regions in the alpha band become hypersynchronous in patients with progressive MCI who converted to Alzheimer’s disease (López et al., 2014a). The increased functional connectivity in the frontal areas was also reported using resting state functional MRI in individuals with amnestic MCI (Qi et al., 2010) and in amyloid-β-positive CN elderly individuals (Mormino et al.,...
Figure 4 Multiple regression analysis between the power marker values and regional grey matter volume (A–C) or regional glucose metabolism (D), adjusting for the effects of age. (A) Results of multiple regression analysis using whole-brain voxel-based morphometry (VBM) for power markers that represent the main effects of clinical category in all subjects (n = 66). Left: Delta power at 2.5 Hz in the posterior part of the brain. Right: Theta power at 4.5 Hz in the global brain. Regions in which the grey matter volumes showed significant negative correlations (FWE-corrected P < 0.05 at a height threshold of P = 0.001, adjusted for the effects of age) were visualized. (B) Results of VBM for the power markers that represent the effects of clinical category within the amyloid-β-negative groups (CNn and MCIp, n = 28). Left: Delta power at 2.5 Hz in the occipitotemporal areas (FWE-corrected P < 0.05 at a height threshold of P = 0.001). Right: Theta power at 4.5 Hz in the occipitotemporal areas (FWE-corrected P < 0.05 at a height threshold of P = 0.005). (C) Results of VBM analyses for power markers that represent the effects of clinical category within the amyloid-β-positive groups (CNp and MCIp, n = 38) as 3.5-Hz delta power in the medial prefrontal areas (FWE-corrected P < 0.05 at a height threshold of P = 0.005). (D) Results of multiple regression analysis of FDG-PET images in the amyloid-β-positive groups (CNp and MCIp, n = 38) for the same power marker as C. Statistically significant clusters in which regional glucose metabolism showed significant negative correlations with the power marker values are visualized (FWE-corrected P < 0.05 at a height threshold of P = 0.001). sMRI = structural MRI.
2011; Lim et al., 2014a; Jones et al., 2015). Although the methodological approach of our study is different from these earlier studies (power spectrum versus functional connectivity), common pathophysiological mechanisms may exist that are related to the frontal functional upregulation. We consider two possible explanations for this. The first hypothesis is a compensatory mechanism, which was also proposed in previous studies (Qi et al., 2010; Mormino et al., 2011; Lim et al., 2014a; Jones et al., 2015). Our results showed a stronger topographical association with a dose-dependent effect between the prefrontal alpha augmentation and prefrontal amyloid-β burden in the CN groups than in the MCI groups and may support this compensatory hypothesis. This is because a sufficient level of compensation is needed to maintain normal cognitive function in the preclinical stages of Alzheimer’s disease, whereas this compensation probably becomes insufficient in the prodromal Alzheimer’s disease stage. The second hypothesis is abnormal hyperexcitability related to amyloid-β deposition. Palop and Mucke (2010) reported a strong influence of amyloid-β in the destabilization of cortical network activity. Busche et al. (2008) demonstrated that clusters of neurons near amyloid plaques become hyperactive, and suggested that this hyperactivity is caused by a relative decrease in synaptic inhibition. This finding was reinforced by a histological study by García-Marin and coworkers (2009), who showed diminished GABAergic terminals in the vicinity of amyloid plaques. These previous studies may support the hyperexcitability hypothesis.

In our study, significant relationships were found between the prefrontal alpha power and local amyloid-β deposition only in group-wise comparisons, but not in the two-way analysis, although the latter revealed a significant main effect of amyloid-β deposition. We consider that this could be due to the differences between the CN and MCI groups in the peak alpha power frequency that showed an amyloid effect. In the group-wise analyses, the peak alpha power frequencies were 9 Hz and 10.5 Hz, respectively, whereas the peak frequency showing the main effect was 9.5 Hz in the two-way analysis. This may indicate that the correlations between the prefrontal alpha power and local amyloid-β deposition were significant only around the peak frequency of alpha power in each clinical category.

The power markers that represented effects of amyloid-β deposition did not show a significant correlation with any of the cognitive measures. This was in line with a previous report (Jack et al., 2009) suggesting that clinical symptoms are not coupled with amyloid-β deposition. In general, downstream topographical markers such as regional glucose hypometabolism measured by FDG-PET and medial temporal atrophy assessed by structural MRI are not considered specific to amyloid-β pathology, especially in the preclinical stage of Alzheimer’s disease (Dubois et al., 2016). Therefore, potential amyloid-β-related biomarker information may be one of the unique features of the MEG markers.

Effects of clinical status on regional power spectra

In the two-way analyses, the main effects of the clinical status were represented as widespread power augmentation within the low-frequency bands (delta and theta) in the MCI groups compared with the CN groups. The power values were significantly correlated with cognitive decline and hippocampal atrophy. In addition, the MCI groups also showed reduced alpha power in the prefrontal areas. These findings were similar to typical spectral patterns observed in previous reports that compared MCI and CN groups (Babiloni et al., 2006; Fernández et al., 2006b, 2013; Rossini et al., 2007; Stam, 2010; Lizio et al., 2011; López et al., 2016). However, when subjects were further segregated based on amyloid-β positivity, group-wise comparisons revealed additional important findings that are crucial for understanding the relevance of regional spectral patterns to their pathophysiological backgrounds.

The group-wise comparison within the amyloid-β-negative groups (MCIn versus CNn) demonstrated similar findings with the two-way comparison. The MCIn group showed increased delta and theta power in rather widespread areas and decreased alpha power in the prefrontal region compared with the CNn group. This is important because such power spectral features were previously considered to be changes related to the progression of Alzheimer’s disease (Fernández et al., 2006a; Rossini et al., 2007). However, the results indicated that these changes are not specific to Alzheimer’s disease and can be observed without Alzheimer’s disease pathology (i.e. amyloid-β deposition). A further important finding was that the power marker values, especially the delta and theta power increase in the posterior brain regions, were significantly correlated with cortical atrophy, specifically in the hippocampus. Recently, the suspected non-Alzheimer disease pathophysiology (SNAP) concept was developed, which suggests the presence of a significant amyloid-β-negative population with biomarker evidence of neurodegeneration (Jack et al., 2016). The prevalence of SNAP is considered to be around 25% in individuals with MCI and CN (Vos et al., 2015; Burnham et al., 2016; Mormino et al., 2016). In fact, 4/25 and 5/11 subjects in the CNn and MCIn groups, respectively (in total 9/36 = 25%), had VSRAD scores >1.0 in our sample, which matches the reported prevalence. Although SNAP represents a heterogeneous status with different pathological aetiologies (Jack et al., 2016; Mormino et al., 2016), delta and theta power augmentation in posterior brain regions may be associated with disease progression within a particular SNAP status.

The results of the group-wise comparison within the amyloid-β-positive groups (MCIp versus CNp) provided even more important information, because they highlighted the regional spectral pattern related to disease progression...
within the Alzheimer’s disease continuum. They were expressed as delta power augmentation in the medial prefrontal and occipital regions in the MCIp group compared to the CNp group. Both markers were significantly correlated with cognitive scores. In particular, the medial prefrontal delta power appeared to be an important MEG marker, because it was significantly correlated with other imaging markers that serve as surrogates for disease progression, including cortical atrophy in the entorhinal cortex and Alzheimer’s disease-like regional glucose hypometabolism. This finding is in line with our previous report suggesting that delta activity in the anterior and occipital brain regions is associated with disease progression (Fernández et al., 2013; López et al., 2016). These results suggest that medial prefrontal power augmentation is coupled with neurodegeneration, and is expected to be useful for monitoring disease progression in the preclinical and prodromal stages of Alzheimer’s disease.

**Potential clinical utility of the MEG power markers**

To estimate the potential clinical utility of the MEG power markers that showed significant group effects of amyloid-β deposition, we evaluated performances to distinguish between amyloid-β-positive and amyloid-β-negative individuals using ROC analyses. Results demonstrated that within the CN groups, the prefrontal alpha power at 10.5 Hz could predict amyloid-β positivity with an area under the curve (AUC) of 0.788 and accuracy of 0.763 (Supplementary Fig. 3A and Supplementary Table 2). Within the MCI groups, the prefrontal alpha power at 9 Hz showed performance with an AUC of 0.866 and

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**Figure 5** A schematic summarizing the main findings. The left and right red arrows and their connected boxes demonstrate the characteristics of the MEG power markers that represented the effects of amyloid-β deposition within the CN groups and within the MCI groups, respectively. The upper and lower red arrows and their connected boxes demonstrate the characteristics of the MEG power markers that represented the effects of clinical category within the amyloid-β-positive groups and within the amyloid-β-negative groups, respectively. The arrows with the gradation colours indicate the directions where the relative power increases (not indicating clinical transition).
MEG signatures of Alzheimer’s disease

BRAIN 2018: 141: 1470–1485 | 1483

accuracy of 0.786 (Supplementary Fig. 3B and Supplementary Table 3).

To identify individuals who showed medial temporal atrophy, we also used ROC analyses to estimate the performances of power markers that showed significant correlations with the grey matter volume in the medial temporal areas. For the analyses, the degree of medial temporal atrophy was dichotomized using the VSRAD scores with a cut-off value of 1.0 (Supplementary Fig. 1). Values lower than the cut-off point suggested that the medial temporal regions were not atrophic. Among the power markers representing the main effects of clinical status, the global theta power augmentation at 4.5 Hz demonstrated the highest performance with an AUC of 0.833 and accuracy of 0.773 (Supplementary Fig. 4A and Supplementary Table 4). For the power markers representing group effects within the amyloid-β-negative groups, the occipitotemporal theta power showed the highest performance with an AUC of 0.831 and accuracy of 0.806 (Supplementary Fig. 4B and Supplementary Table 4). The power marker representing the within-amyloid-β-positive group effects as delta (3.5 Hz) power augmentation in the prefrontal areas showed an AUC of 0.880 and accuracy of 0.867 (Supplementary Fig. 4C and Supplementary Table 4).

However, because these analyses, especially for the amyloid-β markers, could be circular, these values should be only interpreted as references, and validation in an independent dataset is required.

**Limitations of this study**

The present study is limited because of the relatively small sample size and the lack of follow-up information. Validation studies should be carried out with larger sample sizes, preferably coupled with follow-up information. Tau marker information was also not available, which also limits the scope of this study. Combining our methods with direct tau markers, such as CSF or tau PET imaging markers, may deepen our understanding of the association between MEG signatures and their pathophysiological implications.

**Conclusion**

This investigation demonstrated that the regional spectral patterns of resting state MEG activity in MCI and CN subjects conveyed complex information derived from different pathophysiological backgrounds. By incorporating the biomarker information for amyloid-β deposition and neurodegeneration, complex MEG signatures were successfully revealed as summarized in Fig. 5. These findings suggest that MEG potentially offers the following biomarker information: (i) the power augmentation in the alpha band in the medial prefrontal regions is a surrogate marker for amyloid-β pathology both in the CN and MCI groups, and the topographical association with the local amyloid-β burden is stronger in the CN group than the MCI group; (ii) the delta power increase in the medial frontal region is a surrogate marker for disease progression within the Alzheimer’s disease continuum, and is associated with downstream changes, including cortical atrophy in the entorhinal cortex, and Alzheimer’s disease-like regional glucose hypometabolism; and (iii) delta and theta power augmentation in posterior brain regions is a surrogate marker for hippocampal atrophy and general cognitive decline, and the power changes can be observed without amyloid-β pathology, indicating that the signature is not specific to Alzheimer’s disease. These MEG signatures are also expected to deepen the understanding of the pathophysiological processes of disease progression in predementia stages of Alzheimer’s disease.

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**Supplementary material**

Supplementary material is available at Brain online.

**References**


